

Catalytic Environmentally Friendly Protocol for Achmatowicz Rearrangement

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Supporting Information

OH M-CPBA or NBS
$$R_2$$
 OH R_1 R_2 OH R_1 R_2 OH R_2 OH R_3 OF R_4 OF R_4 OF R_4 OF R_5 OF R_6 OF R_7 OF R_8 OF R_8 OF R_8 OF R_8 OF R_8 OF R_9 OF

ABSTRACT: The increasing interest in Achmatowicz rearrangement in organic synthesis calls for a more environmentally friendly protocol since the most popular oxidants m-CPBA and NBS produced stoichiometric organic side product (mchlorobenzoic acid or succinimide). Mechanism-guided analysis enables us to develop a new catalytic method (Oxone/KBr) for AchR in excellent yield with K₂SO₄ as the only side product, which greatly facilitates the purification. This protocol was integrated with other transformations, leading to a rapid access to the highly functionalized dihydropyranones.

chmatowicz rearrangement, an oxidative ring-expansion A chmatowicz rearrangement, and carried furfuryl alcohols to densely functionalized dihydropyranone acetals, has received increasing interest in organic synthesis.² As a powerful and versatile synthetic tool for the preparation of tetrahydropyrans, dihydropyranones, oxidopyrylium, δ -lactones, and pyranoses, etc., AchR could be performed with various oxidation methods,³ including Br₂/MeOH,¹ N-bromosuccinimide (NBS),4 in situ generated dimethyldioxirane (DMDO),5 mchloroperoxybenzoic acid (m-CPBA),6 magnesium monoperoxypthalate, metal-base oxidant (PCC, VO(acac)₂/TBHP, 9 titanium(IV) silicalite/H₂O₂¹⁰), phenyliodine(III) diacetate (PIDA), ¹¹ photolytic oxidation $(O_2/h\nu)$, ¹² electrochemical oxidation, ¹³ and enzymatic transformations. ¹⁴ Among these methods, NBS and m-CPBA are the most widely used oxidants for their simple operation in practice, tolerance of many functional groups, and reliably high yield in most cases. However, the major drawback of these two protocols is the generation of stoichiometric organic side product (mchlorobenzoic acid or succinimide), which usually requires immediate purification by column chromatography. Catalytic variants of these two primary methods are not available, which fact prompted us to develop a green, catalytic protocol for AchR with an ultimate goal of no generation of the direct organic side products derived from both the oxidant and the catalyst employed.

Mechanistic consideration of NBS-mediated AchR guided us to explore low-cost, nontoxic, environmentally friendly Oxone¹⁵ (2KHSO₅-KHSO₄-K₂SO₄) as the oxidant coupled with an inorganic halide salt as the catalyst (Figure 1). We conceived that the oxidation of an alkali bromide with Oxone

Figure 1. Achmatowicz rearrangement under classical conditions and our catalytic protocol with possible mechanism.

might generate an active transient brominating agent ([Br⁺] such as HOBr or Br₂), ¹⁶ which would promote AchR of furfuryl alcohols in a similar manner as NBS or Br₂.⁴ The ring

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Table 1. Selected Conditions for Catalytic Achmatowicz Rearragement with Oxone/Halides

entry	oxidant/halide	additive (equiv)	solvent	time	yield (%
1	Oxone/NH ₄ Br	NaHCO ₃ (2)/NaOAc (1)	THF/H_2O (4/1)	30 min	62
2	Oxone/LiBr	NaHCO ₃ (2)/NaOAc (1)	THF/ H_2O (4/1)	30 min	80
3	Oxone/NaBr	NaHCO ₃ (2)/NaOAc (1)	THF/H_2O (4/1)	30 min	82
4	Oxone/CaBr ₂	NaHCO ₃ (2)/NaOAc (1)	THF/ H_2O (4/1)	30 min	77
5	Oxone/KBr	NaHCO ₃ (2)/NaOAc (1)	THF/ H_2O (4/1)	30 min	82
6	Oxone/KBr	NaHCO ₃ (2)/NaOAc (1)	$MeOH/H_2O$ (1/1)	30 min	nd
7	Oxone/KBr	NaHCO ₃ (2)/NaOAc (1)	DCM/H_2O (4/1)	48 h	<5
8	Oxone/KBr	NaHCO ₃ (2)/NaOAc (1)	$MeCN/H_2O$ (20/1)	30 min	79
9	Oxone/KBr		THF/H_2O (4/1)	30 min	49
10	Oxone/KBr	NaHCO ₃ (2)	THF/ H_2O (4/1)	30 min	73
11	Oxone/KBr	$NaHCO_3$ (1)	THF/ H_2O (4/1)	30 min	83
12	Oxone/KBr	$NaHCO_3$ (0.5)	THF/ H_2O (4/1)	30 min	93
13	Oxone/KBr	NaHCO ₃ (0.25)	THF/H_2O (4/1)	30 min	92
14	Oxone/KBr	$NaHCO_3$ (0.1)	THF/ H_2O (4/1)	30 min	86
15	Oxone/NaCl	$NaHCO_3$ (0.5)	THF/ H_2O (4/1)	30 h	17
16	Oxone/NaI	$NaHCO_3$ (0.5)	THF/ H_2O (4/1)	30 h	<5
17	Oxone/NBS	$NaHCO_3$ (0.5)	THF/ H_2O (4/1)	30 min	71
18	Oxone/m-CPBA	NaHCO ₃ (0.5)	THF/ H_2O (4/1)	48 h	35
19	Oxone	$NaHCO_3$ (0.5)	THF/H_2O (4/1)	24 h	<20
20	H_2O_2/KBr	NaHCO ₃ (0.5)	THF/ H_2O (4/1)	48 h	<5
21	H ₂ O ₂ /NBS	$NaHCO_3$ (0.5)	THF/ H_2O (4/1)	48 h	<5

expansion (ring opening and subsequent ring closure) would produce the dihydropyranone acetal with generation of K₂SO₄ (potassium used as the alkali counterion) as the only side product and release of the catalytic bromide, which would be oxidized again by Oxone for the subsequent catalytic cycles. If this hypothetic mechanism works, a truly green, catalytic, and practical protocol for AchR could be developed. However, at the beginning stage of our investigations we were very concerned about the potentially competing (1) halogenation of the electron-rich furan of type 1, (2) alcohol oxidation of the furfuryl alcohol, and (3) dihalogenation of the resulting enone functionality of AchR products because the combination of Oxone and halide $(Oxone/MX)^{16}$ has been widely used in oxidation reactions such as halogenation of arenes, dihalogenation oxidation, a lakenes, α -halogenation of ketones, alcohol oxidation, benzylic oxidation, and halolactonization of alkenoic acids/amides. Nevertheless, the chance of successful AchR with Oxone and halide exists if AchR via our hypothetic mechanism precedes oxidation or halogenation reactions.

To verify our mechanism-guided hypothesis, we carried out AchR of 1a with Oxone and different halides under various conditions (Table 1). To our delight, a combination of catalytic amount of bromide (5 mol %) and stoichiometric Oxone (1.2 equiv)²³ was found to be remarkably efficient (49–93% yield) for AchR of 1a when a 4/1 mixture of THF and H₂O was used as the solvent (entries 1-5 and 9-14). It was noteworthy that no furan bromination or alcohol oxidation was observed in the NMR spectra of the crude reaction mixture. As shown in Table 1, the change in counterion (NH₄⁺, Li⁺, Na⁺, K⁺, Ca²⁺, etc.) has little effect on the yield and reaction rate (entries 1-5). However, the reaction medium played a crucial role: MeOH/ H₂O (entry 6) and CH₂Cl₂/H₂O (entry 7) were not suitable for AchR with Oxone/bromide, while MeCN/H₂O (20/1)

(entry 8) was a comparable solvent system (79%). On the other hand, the acidity of the reaction medium was found to be a minor factor on the yield (entries 9-14): addition of 0.25-0.5 equiv of NaHCO3 gave the best yield (92-93%) of 2a within 30 min (entries 12 and 13). It should be noted that this optimized protocol was developed along with the following control experiments: (i) substitution of the bromide with chloride or iodide led to lower or no conversion (entries 15 and 16); (ii) substitution of the bromide with catalytic amount of NBS (entry 17) or m-CPBA (entry 18) reduced the efficiency of AchR, providing 2a with substantially lower yields (71% and 35%, respectively); (iii) no reaction was observed in the absence of the catalytic bromide (entry 19); and (iv) replacement of Oxone with H₂O₂ (or tBuOOH, PhI(OAc)₂, etc.) using catalytic bromide (entry 20) or NBS (entry 21) terminated the AchR.

To evaluate the advantages and disadvantages of our new catalytic protocol under the optimized conditions, we examined the substrate scope using both our new protocol and the conventional NBS- and *m*-CPBA-promoted methods (Table 2). In most cases, our catalytic Oxone/KBr could promote the clean AchR in better or competitive yield (74-93% yield, method A) without a need of purification by flash column chromatography when compared to NBS (method B) or m-CPBA (method C). Importantly, the Oxone/KBr system tolerated various functional groups including silyl ether (2b and 2q, 83-85% yield), ester (2c, 78% yield), alkene (2d and 2j, 75-83% yield, respectively), ketone (2e, 88% yield), Weinreb amide (2i, 90% yield), Evans chiral oxazolidinone (2k, 90% yield), and electron-rich arenes (2m-o and 2s, 74-91% yield). There were no potentially competing side reactions including arene bromination, ketone α -bromination, alkene dibromination, and alcohol oxidation, all of which have been The Journal of Organic Chemistry

Table 2. Substrate Scope and Comparison with NBS and m-CPBA^a

Method A: KBr (5 mol%), Oxone (1.2 eq), NaHCO $_3$ (0.5 eq) in THF/H $_2$ O (4/1) at 0°C **Method B**: NBS (1.2 eq), NaHCO $_3$ (2 eq), NaOAc (1 eq) in THF/H $_2$ O (4/1) at 0°C **Method C**: m-CPBA (1.2 eq) in DCM at 0°C

^aIsolated yield. ^bReaction was carried out with KBr (20 mol %), Oxone (1.2 equiv), and NaHCO₃ (0.5 equiv) in a mixture of THF and H₂O (4/1) at 0 °C. ^cReaction was carried out with KBr (5 mol %), Oxone (1.2 equiv), and NaHCO₃ (2 equiv) in a mixture of THF and H₂O (4/1) at 0 °C.

reported in the reactions using a combination of Oxone and stoichiometric bromide. In particular, the benzyl alcohols, substrates that readily undergo oxidation with Oxone/bromide to aldehydes, could be used in the AchR using our catalytic Oxone/KBr combination reagent (2f and 2s, 80% and 93% yield, respectively). Primary and tertiary furyl alcohols were also excellent substrates for our catalytic AchR to afford the desired products (2g, 2h, and 2r) in excellent yields (85–87%). Remarkably, the aza-AchR²⁴ of furyl sulfonamide using Oxone/KBr proceeded more efficiently (86% yield) than the use of NBS (69% yield) and *m*-CPBA (74% yield). Notably, compounds (2l-o and 2q) were the key intermediates for total synthesis of musellarins²⁵ and uprolides.²⁶

The practicality of this new catalytic protocol was further examined for scalability and stereochemistry integrity (Scheme 1). Toward this end, the optically active furfuryl alcohol 1b (98% ee) was subjected to our standard conditions using Oxone/KBr for AchR on a 6.0 g scale (20 mmol) and delivered 2b in 83% yield, which could be used in the subsequent reactions without flash column chromatography. A three-step functionalization of 2b, acetylation, γ -deoxygenation, and Heck–Matsuda coupling, provided 3 for determination of the optical purity, which revealed no loss of the ee value (97% ee, the ee value was determined by chiral HPLC). These results

Scheme 1. Achmatowicz Rearrangement of Optical Active Furfuryl Alcohol 1b on a Multigram Scale

clearly suggested the laboratory scalability and stereochemistry integrity of our new catalytic protocol, which will become the first choice among the various methods for AchR. Note: caution should be taken for a large-scale reaction since THF can react with bromine under vigorous gas-producing reactions possibly via photocatalysis, and light effects should be included in safety reviews before any large-scale work is undertaken.

The absence of organic side products when using Oxone/KBr offered a great opportunity to us for investigations on AchR-participating one-pot sequential reactions (Scheme 2). Two illustrative examples are shown in Scheme 2. Treatment of furfuryl diol $1 u^{27}$ with Oxone (1.2 equiv) and KBr (5 mol %) in a mixture of MeCN and H_2O (20/1) at 0 °C for 30 min gave the AchR product 2 u, which upon treatment of CSA (1 equiv) in the same reaction vessel underwent efficient spiroketalization to provide 4 in 71% overall yield. Similarly, the one-pot AchR/

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Scheme 2. AchR-Participating One-Pot Sequential Reactions

bromination was practically efficient: additions¹⁷ of KBr (1.0 equiv) and Oxone (1.0 equiv) to the crude AchR product 2m gave the monobrominated product 5 in 85% yield.

Since column purification is usually required for NBS and *m*-CPBA methods to remove the organic side products (succinimide and p-chlorobenzoic acid) that might prevent subsequent transformations, our catalytic protocol was highly efficient and did not produce any organic side products derived from catalyst (KBr) and oxidant (Oxone). In order to demonstrate such an operational advantage, a number of classical transformations were performed using the nonpurified 2a from Oxone/KBr-mediated AchR (Scheme 3). The crude

Scheme 3. Use of the Crude AchR Products for Subsequent **Transformations**

AchR product 2a (obtained by simple extraction, drying over MgSO₄, and evaporation of the organic solvents) smoothly underwent γ -deoxygenation $(2a \rightarrow 6)$, 25 acetylation $(2a \rightarrow 7a)$, 28 carbonate formation $(2a \rightarrow 7b)$, 29 and Jones oxidation $(2a \rightarrow 8)^{30}$ in good to excellent yield. It was noted that Kishi³¹ reduction $(2a \rightarrow 9)$ was not successful due to over-reduction and other unknown side reactions. However, the 2,6disubstituted dihydropyranone acetal 2q could undergo efficient Kishi reduction to provide cis-2,6-disubstituted dihydropyranone 10, which was a key intermediate in our total synthesis of uprolides. 26 The successful implementation of these transformations greatly expanded the utility of this protocol in organic synthesis.2

In summary, a mechanism-guided analysis enabled us to develop a new, practical, catalytic protocol for Achmatowicz rearrangement, featuring (1) the use of environmentally friendly, nontoxic, easy to handle, cheap, and stable Oxone as the terminal oxidant, (2) employment of KBr as the catalyst, (3) no organic wastes derived from oxidant and catalyst, and (4) no need for column chromatography for purification. The efficiency of this protocol was fully demonstrated in 20 examples and compared with the classical methods using NBS and m-CPBA as the oxidant. In addition, the Oxone/KBr protocol for Achmatowicz rearrangement was integrated with other subsequent transformations, leading to a rapid access to highly functionalized dihydropyranones through sequential reactions and/or subsequent functionalization of the crude AchR products.

■ EXPERIMENTAL SECTION

General Experimental Methods. Reactions were carried out in oven- or flame-dried glassware under a nitrogen atmosphere, unless otherwise noted. Tetrahydrofuran (THF) was freshly distilled before use from sodium using benzophenone as indicator. Dichloromethane (DCM) was freshly distilled before use from calcium hydride (CaH₂). All other anhydrous solvents were dried over 3 or 4 Å molecular sieves. Solvents used in workup, extraction, and column chromatography were used as received from commercial suppliers without prior purification. Reactions were magnetically stirred and monitored by thin-layer chromatography (TLC, 0.25 mm) on precoated silica gel plates. Flash chromatography was performed with silica gel 60 (particle size 0.040-0.062 mm). Infrared spectra were measured with neat sample. ¹H and ¹³C NMR spectra were recorded on a 400 MHz spectrometer (400 MHz for ¹H, 100 MHz for ¹³C). Chemical shifts are reported in parts per million (ppm) as values relative to the internal chloroform (7.26 ppm for ¹H and 77.16 ppm for ¹³C), benzene (7.16 ppm for ¹H and 128.06 ppm for ¹³C), methanol (3.31 ppm for ¹H and 49.00 ppm for ¹³C), and acetone (2.09 ppm for ¹H and 30.60 ppm for ¹³C). Abbreviations for signal coupling are as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet. Highresolution mass spectra were measured using TOF as the analyzer.

Preparation of 1-(Furan-2-yl)-3-(4-((triisopropylsilyl)oxy)phenyl)propan-1-ol (1n). To a stirred solution of 3-(4hydroxyphenyl)propionaldehyde (1.52 g, 10 mmol) in anhydrous DCM (30 mL) were added imidazole (1.71 g, 25.1 mmol) and triisopropoylsilyl chloride (TIPSCl, 2.31 g, 12 mmol) at 0 °C. After completion of the addition, the reaction mixture was allowed to warm to room temperature and stirred overnight. The reaction was quenched by addition of water (10 mL). The organic fractions were collected, and the aqueous phase was extracted with CH_2Cl_2 (3 × 20 mL). The combined organic fractions were washed with saturated aqueous NH₄Cl solution and brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The crude protection product was used for the next step without further purification. To a stirred solution of furan (2.72 g, 40 mmol) in anhydrous THF (50 mL) was added n-BuLi (1.6 M in cyclohexane, 12.5 mL, 20 mmol) slowly at -78 °C. After completion of the addition, the reaction mixture was allowed to warm to -20 °C for 1 h. The crude product above was dissolved in THF (10 mL) and then added slowly to the lithiated furan solution at -78 °C. After completion of the addition, the reaction mixture was monitored by TLC. The reaction was quenched by addition of saturated aqueous NH₄Cl and extracted with EtOAc. The combined organic fractions were washed with brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 5:1) to give the substituted furfuryl alcohol 1n (3.06 g, 8.12 mmol) as a yellowish oil in 81% yield over two steps. IR (neat, cm⁻¹): 3398, 2947, 2862, 1510, 1292, 1231, 1140, 997, 880, 678. ¹H NMR (400 MHz, CDCl₃) δ : 7.37 (d, J = 1.9 Hz, 1H), 7.05 (d, J = 8.1 Hz, 2H), 6.81 (d, J = 8.3 Hz, 2H), 6.33 (t, J = 2.4 Hz, 1H), 6.23 (d, J = 3.2 Hz, 1H), 4.66 (t, J = 6.8 Hz, 1H), 2.76-2.57 (m, 2H), 2.14 (q, J = 7.5 Hz, 2H), 1.31-1.21 (m, 3H), 1.13-1.06 (m, 21H). 13 C NMR (100 MHz, CDCl₃) δ : 156.8, 154.3, 142.0, 133.9, 129.4, 119.9, 110.2, 106.1, 67.1, 37.3, 31.0, 18.1 (6 × C), 12.8 (3 × C). HRMS (TOF, CI⁺): m/z calcd for $C_{22}H_{34}O_3Si$ [M]⁺ 374.2277, found 374.2266.

General Procedure A: Achmatowicz Rearrangement Using Oxone and Catalytic KBr. To a stirred solution of the furfuryl alcohol (0.5 mmol) in THF (4 mL) and $\rm H_2O$ (1 mL) were added KBr (5.9 mg, 0.025 mmol), NaHCO $_3$ (22 mg, 0.25 mmol), and Oxone (0.37 g, 0.6 mmol) at 0 °C. After completion of the addition, the reaction was allowed to stir at 0 °C for 30 min. The reaction was then quenched by addition of saturated aqueous NaHCO $_3$ (10 mL) and EtOAc (3 × 10 mL). The combined organic fractions were washed with brine, dried over Na $_2$ SO $_4$, and concentrated under reduced pressure.

General Procedure B: Achmatowicz Rearrangement Using Stoichiometric NBS. To a stirred solution of the furfuryl alcohol (0.5 mmol) in THF (4 mL) and $\rm H_2O$ (1 mL) were added NaHCO₃ (85 mg, 1 mmol), NaOAc (40 mg, 0.5 mmol), and N-bromosuccinimide (NBS, 90 mg, 0.5 mmol) at 0 °C. After completion of the addition, the reaction was allowed to stir at 0 °C for 30 min. The reaction was then quenched by addition of saturated aqueous NaHCO₃ (10 mL) and Na₂S₂O₃ (10 mL) and extracted with EtOAc (3 × 10 mL). The combined organic fractions were washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (EtOAc/hexane = 1/4–1/2) to afford the desired product.

General Procedure C: Achmatowicz Rearrangement Using Stoichiometric m-CPBA. To a stirred solution of the furfuryl alcohol (0.5 mmol) in DCM (4 mL) was added m-chloroperbenzoic acid (m-CPBA, 77%, 0.17 g, 0.75 mmol) at 0 °C. After completion of the addition, the reaction was allowed to stir at 0 °C for 30 min—4 h. The reaction was then quenched by addition of saturated aqueous NaHCO₃ (10 mL) and Na₂S₂O₃ (10 mL) and extracted with CH₂Cl₂ (3 × 10 mL). The combined organic fractions were washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (EtOAc/hexane = 1/4–1/2) to afford the desired product.

6-Hydroxy-2-isopropyl-2H-pyran-3(6H)-one (2a). ²⁵ Yellowish oil (dr 3:2; method A, 72 mg, 92%; method B, 62 mg, 79%; method C, 60 mg, 77%). Major diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 6.93–6.87 (m, 1H), 6.13–6.07 (m, 1H), 5.65 (d, J = 4.0 Hz, 1H), 4.39 (dd, J = 3.2, 1.1 Hz, 1H), 2.46–2.38 (m, 1H), 1.04–1.00 (m, 3H), 0.86 (dd, J = 6.9, 1.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 197.1, 144.7, 128.1, 87.7, 78.5, 28.7, 19.1, 16.4. Minor diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 6.93–6.87 (m, 1H), 6.13–6.07 (m, 1H), 5.62 (t, J = 4.0 Hz, 1H), 3.93–3.86 (m, 1H), 2.46–2.38 (m, 1H), 1.04–1.00 (m, 3H), 0.91 (d, J = 6.8 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 196.6, 148.5, 129.5, 91.3, 83.2, 29.0, 19.2, 16.6.

Gram-Scale Reaction of 6-Hydroxy-2-(2-((triisopropylsilyl)oxy)-ethyl)-2H-pyran-3(6H)-one (1b). To a stirred solution of furfuryl alcohol (+)-1b (6.01 g, 20.1 mmol) in THF (40 mL) and water (10 mL) at 0 °C were added NaHCO $_3$ (0.85 g, 10.07 mmol), KBr (0.12 g, and Oxone (14.8 g, 24.1 mmol). The reaction mixture was stirred at 0 °C for 30 min. The reaction was then quenched by addition of saturated aqueous NaHCO $_3$ (100 mL) and extracted with EtOAc (3 \times 30 mL). The combined organic fractions were washed with brine, dried over Na $_2$ SO $_4$, and concentrated under reduced pressure. The crude product 2b (5.23 g, 16.7 mmol) was obtained in 83% yield and used for next step without further purification.

6-Hydroxy-2-(2-((triisopropylsilyl)oxy)ethyl)-2H-pyran-3(6H)-one (**2b**). Yellowish oil (dr 2:1; method A, 134 mg, 85%; method B, 123 mg, 78%; method C, 105 mg, 67%). $\left[\alpha_D^{20}\right]$ = +53.2 (c 1.0, MeOH). IR (neat, cm⁻¹): 3385, 2968, 2942, 2889, 2864, 1689, 1464, 1267, 1135, 1010. Major diastereomer: ¹H NMR (400 MHz, CDCl₃) δ : 6.93–6.86 (m, 1H), 6.15–6.08 (m, 1H), 5.62 (t, J = 3.3 Hz, 1H), 4.78 (dd, J = 8.4, 3.9 Hz, 1H), 3.98–3.79 (m, 2H), 2.28–2.20 (m, 1H), 1.94–1.78 (m, 1H), 1.14–0.98 (m, 21H). ¹³C NMR (100 MHz, CDCl₃) δ : 196.9, 144.3, 127.7, 87.7, 71.0, 59.0, 33.1, 18.1 (δ × C), 12.1 (δ × C). Minor

diastereomer: 1 H NMR (400 MHz, CDCl₃) δ : 6.93–6.86 (m, 1H), 6.15–6.08 (m, 1H), 5.62 (t, J = 3.3 Hz, 1H), 4.35 (dd, J = 9.0, 3.7 Hz, 1H), 3.89–3.81 (m, 2H), 2.28–2.20 (m, 1H), 1.96–1.78 (m, 1H), 1.14–0.98 (m, 21H). 13 C NMR (100 MHz, CDCl₃) δ : 196.6, 147.8, 128.8, 90.9, 75.7, 59.0, 33.9, 18.1 (6 × C), 12.1 (3 × C). HRMS (Cl⁺): m/z calcd for $C_{16}H_{31}O_4$ Si [M + H]⁺ 315.1986, found 315.1993.

Ethyl 2-(6-Hydroxy-3-oxo-3,6-dihydro-2H-pyran-2-yl)acetate (2c). Yellowish oil (dr 5:2; method A, 78 mg, 78%; method B, 75 mg, 75%; method C, 82 mg, 82%). Major diastereomer. H NMR (400 MHz, CDCl₃) δ : 6.92 (dd, J = 10.3, 3.5 Hz, 1H), 6.14 (d, J = 10.2 Hz, 1H), 5.63 (d, J = 3.5 Hz, 1H), 5.02 (dd, J = 7.7, 3.8 Hz, 1H), 4.16 (qd, J = 7.1, 3.8 Hz, 2H), 3.00 (dt, J = 16.8, 3.6 Hz, 1H), 2.87–2.67 (m, 1H), 1.26 (td, J = 7.1, 2.8 Hz, 3H). 13 C NMR (100 MHz, CDCl₃) δ : 195.0, 171.1, 144.7, 127.3, 87.9, 70.9, 61.2, 35.4, 14.2. Minor diastereomer. H NMR (400 MHz, CDCl₃) δ : 6.96 (dd, J = 10.4, 1.5 Hz, 1H), 6.18 (dd, J = 10.4, 1.5 Hz, 1H), 5.71 (d, J = 1.7 Hz, 1H), 4.57 (ddd, J = 7.9, 3.8, 1.2 Hz, 1H), 4.16 (qd, J = 7.1, 3.8 Hz, 2H), 3.00 (dt, J = 16.8, 3.6 Hz, 1H), 2.87–2.67 (m, 1H), 1.26 (td, J = 7.1, 2.8 Hz, 3H). 13 C NMR (100 MHz, CDCl₃) δ : 194.5, 171.1, 148.4, 128.5, 91.0, 75.5, 61.3, 36.2, 14.2.

2-Allyl-6-hydroxy-2H-pyran-3(6H)-one (2d).³³ Yellowish oil (dr 5:2; method A, 57.8 mg, 75%; method B, 46.2 mg, 60%; method C, 63.2 mg, 82%). Major diastereomer. ¹H NMR (400 MHz, C_6D_6) δ: 6.25 (dd, J=10.3, 3.5 Hz, 1H), 5.95–5.77 (m, 2H), 5.22–5.00 (m, 3H), 4.58 (dd, J=7.7, 3.9 Hz, 1H), 2.80–2.66 (m, 1H), 2.59–2.44 (m, 1H). ¹³C NMR (100 MHz, C_6D_6) δ: 196.0, 145.2, 134.3, 127.1, 117.8, 87.8, 73.9, 34.5. Minor diastereomer. ¹H NMR (400 MHz, C_6D_6) δ: 6.36 (dd, J=10.3, 1.5 Hz, 1H), 5.95–5.77 (m, 2H), 5.22–5.00 (m, 3H), 4.58 (dd, J=7.7, 3.9 Hz, 1H), 2.80–2.66 (m, 1H), 2.59–2.44 (m, 1H). ¹³C NMR (100 MHz, C_6D_6) δ: 195.4, 148.7, 134.2, 127.1, 117.9, 91.2, 78.5, 35.2.

6-Hydroxy-2-(2-oxopropyl)-2H-pyran-3(6H)-one (2e). Yellowish oil (dr 2:1; method A, 75 mg, 88%; method B, 69 mg, 81%; method C, 62 mg, 73%). IR (neat, cm $^{-1}$): 3390, 2970, 2938, 2881, 2858, 1686, 1568 1470, 1275, 1145, 1065, 987. Major diastereomer. 1 H NMR (400 MHz, CDCl₃) δ: 6.90 (dd, J = 10.3, 3.5 Hz, 1H), 6.10 (d, J = 10.3 Hz, 1H), 5.58 (d, J = 3.5 Hz, 1H), 5.03 (dd, J = 7.5, 3.8 Hz, 1H), 3.11 (dd, J = 17.6, 3.7 Hz, 1H), 2.82 (dd, J = 17.6z, 7.5 Hz, 1H), 2.20 (s, 3H). 13 C NMR (100 MHz, CDCl₃) δ: 206.1, 195.8, 144.9, 127.0, 87.8, 70.2, 43.8, 30.5. Minor diastereomer. 1 H NMR (400 MHz, CDCl₃) δ: 6.95 (dd, J = 10.3, 1.4 Hz, 1H), 6.15 (dd, J = 10.3, 1.7 Hz, 1H), 5.70 (d, J = 1.8 Hz, 1H), 4.59 (ddd, J = 7.6, 3.8, 1.4 Hz, 1H), 3.11 (dd, J = 17.6, 3.7 Hz, 1H), 2.90 (dd, J = 17.8, 7.6 Hz, 1H), 2.19 (s, 3H). 13 C NMR (100 MHz, CDCl₃) δ: 206.0, 195.3, 148.8, 128.5, 91.1, 74.7, 44.2, 30.6. HRMS (Cl $^{+}$): m/z calcd for C_8 H₁₁O₄ [M + H] $^{+}$ 171.0652, found 171.0658.

6-Hydroxy-2-phenyl-2H-pyran-3(6H)-one (2f). ³⁴ Yellowish oil (dr 2:1; method A, 76 mg, 80%; method B, 83 mg, 87%; method C, 75 mg, 79%). Major diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 7.37–7.30 (m, 5H), 6.92–6.86 (m, 1H), 6.21–6.14 (m, 1H), 5.61 (d, J = 2.4 Hz, 1H), 5.55 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ: 195.0, 145.4, 135.3, 130.1, 129.2, 128.7, 127.6, 87.9, 77.0. Minor diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 7.37–7.30 (m, 5H), 6.92–6.86 (m, 1H), 6.21–6.14 (m, 1H), 5.65 (s, 1H), 5.01 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ: 194.6, 145.4, 129.5, 129.2, 128.7, 128.5, 128.0, 91.5, 81.1.

6-Hydroxy-2,2-dimethyl-2H-pyran-3(6H)-one (**2g**). ³⁵ Yellowish oil (method A, 62 mg, 87%; method B, 57 mg, 80%; method C, 51.2 mg, 72%). ¹H NMR (400 MHz, CDCl₃) δ : 6.78 (dd, J = 10.3, 3.1 Hz, 1H), 6.09 (dd, J = 10.5, 3.2 Hz, 1H), 5.78 (d, J = 2.9 Hz, 1H), 1.48 (s, 3H), 1.39 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ : 198.9, 143.3, 126.6, 89.2, 79.5, 27.4, 25.0.

2-Hydroxy-1-oxaspiro[5.5]undec-3-en-5-one (2h).³² Yellowish oil (method A, 74 mg, 81%; method B, 71 mg, 78%; method C, 69 mg, 76%). ¹H NMR (400 MHz, CDCl₃) δ : 6.82 (dd, J = 10.2, 2.0 Hz, 1H), 6.01 (d, J = 10.3 Hz, 1H), 5.66 (s, 1H), 4.40 (brs, 1H), 1.92–1.45 (m, 9H), 1.31–1.13 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ : 199.7, 146.0, 126.7, 87.5, 80.7, 33.4, 31.0, 25.2, 21.0, 20.6.

2-(6-Hydroxy-3-oxo-3,6-dihydro-2H-pyran-2-yl)-N-methoxy-N-methylacetamide (2i). Yellowish oil (dr 5:2; method A, 97 mg, 90%; method B, 82 mg, 76%; method C, 91.5 mg, 85%). IR (neat, cm⁻¹):

3395, 2976, 2962, 2920, 2870, 1693, 1665, 1478, 1375, 1277, 1145, 1105, 956. Major diastereomer. 1H NMR (400 MHz, CDCl₃) δ : 6.90 (dd, J = 10.2, 3.5 Hz, 1H), 6.09 (d, J = 10.2 Hz, 1H), 5.58 (d, J = 3.5 Hz, 1H), 5.09 (dd, J = 8.2, 3.4 Hz, 1H), 3.68 (s, 3H), 3.16 (s, 3H), 3.10–3.00 (m, 1H), 2.99–2.85 (m, 1H). 13 C NMR (100 MHz, CDCl₃) δ : 196.1, 171.3, 145.2, 126.9, 87.8, 70.6, 61.4, 33.0. Minor diastereomer. 1 H NMR (400 MHz, CDCl₃) δ : 6.95 (dd, J = 10.3, 1.5 Hz, 1H), 6.15 (dd, J = 10.3, 1.5 Hz, 1H), 5.70 (d, J = 1.7 Hz, 1H), 4.62 (dd, J = 8.0, 3.5 Hz, 1H), 3.68 (s, 3H), 3.16 (s, 3H), 3.10–3.00 (m, 1H), 2.99–2.85 (m, 1H). 13 C NMR (100 MHz, CDCl₃) δ : 195.5, 171.3, 149.0, 128.2, 90.9, 75.0, 61.4, 32.3. HRMS (Cl $^+$): m/z calcd for $C_9H_{14}NO_5$ [M + H] $^+$ 216.0866, found 216.0876.

2-Allyl-6-hydroxy-6-methyl-2H-pyran-3(6H)-one (2j).³⁶ Yellowish oil (dr 6:1; method A, 70 mg, 83%; method B, 57.2 mg, 68%; method C, 66.4 mg, 79%). ¹H NMR (400 MHz, CDCl₃) δ: 6.80 (d, *J* = 10.1 Hz, 1H), 5.98 (d, *J* = 10.1 Hz, 1H), 5.81 (ddt, *J* = 17.1, 10.3, 6.7 Hz, 1H), 5.12 (dd, *J* = 17.2, 1.8 Hz, 1H), 5.04 (dd, *J* = 10.2, 1.8 Hz, 1H), 4.56 (dd, *J* = 7.6, 3.9 Hz, 1H), 3.45 (s, 1H), 2.66 (ddd, *J* = 15.0, 6.1, 4.4 Hz, 1H), 2.41 (dt, *J* = 14.9, 7.3 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ: 196.6, 148.5, 133.8, 126.3, 117.6, 92.9, 74.0, 34.0, 28.8.

(4S)-3-((2S)-2-(6-Hydroxy-3-oxo-3,6-dihydro-2H-pyran-2-yl)-propanoyl)-4-isopropyloxazolidin-2-one (2k). Yellowish oil (dr 5:2; method A, 134 mg, 90%; method B, 126.3 mg, 85%; method C, 116 mg, 78%). [α_D^{20}] = +33.6 (c 1.0, CH₂Cl₂). IR (neat, cm⁻¹): 3390, 2970, 2967, 2930, 2875, 1687, 1658, 1475, 1370, 1272, 1140, 1108, 950. Major diastereomer. ¹H NMR (400 MHz, C₆D₆) δ: 6.15 (dd, J = 10.3, 3.4 Hz, 1H), 5.76 (dt, J = 10.3, 1.8 Hz, 1H), 5.20–5.09 (m, 1H), 4.69 (q, J = 7.5 Hz, 1H), 4.18 (tt, J = 7.7, 3.9 Hz, 1H), 3.55–3.36 (m, 2H), 2.16–2.04 (m, 1H), 1.60–1.49 (m, 3H), 0.56 (d, J = 7.0 Hz, 3H), 0.44 (d, J = 6.9 Hz, 3H). ¹³C NMR (100 MHz, C₆D₆) δ: 196.6, 174.9, 154.2, 148.8, 144.6, 127.0, 87.4, 74.5, 63.3, 58.6, 39.3, 28.9, 17.6, 14.8. HRMS (Cl⁺): m/z calcd for C₁₄H₂₀NO₆ [M + H]⁺ 298.1285, found 298.1279.

6-Hydroxy-2-phenethyl-2H-pyran-3(6H)-one (2I). Yellowish oil (dr 4:3; method A, 96 mg, 88%; method B, 80 mg, 73%; method C, 89.5 mg, 82%). IR (neat, cm $^{-1}$): 3388, 2963, 2935, 2892, 2860, 1683, 1461, 1262, 1130, 1006. Major diastereomer. 1 H NMR (400 MHz, CDCl $_{3}$) δ: 7.34–7.21 (m, 5H), 6.97–6.90 (m, 1H), 6.18–6.12 (m, 1H), 5.69–5.66 (m, 1H), 4.61 (dd, J = 8.3, 3.7 Hz, 1H), 2.88–2.71 (m, 2H), 2.32–2.25 (m, 1H), 2.15–2.03 (m, 1H). 13 C NMR (100 MHz, CDCl $_{3}$) δ: 197.1, 148.4, 145.0, 141.4, 128.5, 127.5, 126.2, 87.7, 73.3, 31.5, 31.1. Minor diastereomer. 1 H NMR (400 MHz, CDCl $_{3}$) δ: 7.34–7.21 (m, 5H), 6.97–6.90 (m, 1H), 6.18–6.12 (m, 1H), 5.69–5.66 (m, 1H), 4.07 (dd, J = 8.8, 4.0 Hz, 1H), 2.88–2.71 (m, 2H), 2.32–2.25 (m, 1H), 2.15–2.03 (m, 1H). 13 C NMR (100 MHz, CDCl $_{3}$) δ: 196.7, 145.0, 141.2, 128.7, 128.6, 126.4, 126.2, 91.0, 77.8, 32.2, 31.1. HRMS (CI $^{+}$): m/z calcd for C $_{13}$ H $_{14}$ O $_{3}$ [M] $^{+}$ 218.0943, found 218.0945.

6-Hydroxy-2-(4-methoxy-3-((triisopropylsilyl)oxy)phenethyl)-2Hpyran-3(6H)-one (2m). Yellowish oil (dr 2:1; method A, 191.4 mg, 91%; method B, 189 mg, 90%; method C, 172.5 mg, 82%). IR (neat, cm⁻¹): 3398, 2961, 2935, 2887, 2864, 1687, 1458, 1260, 1134, 1017. Major diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 6.91-6.86 (m, 1H), 6.75-6.70 (m, 2H), 6.13-6.07 (m, 1H), 5.64-5.60 (m, 1H), 4.52 (dd, J = 8.3, 3.7 Hz, 1H), 3.76 (s, 3H), 2.78–2.54 (m, 2H), 2.23– 2.15 (m, 1H), 2.02-1.90 (m, 1H), 1.33-1.18 (m, 3H), 1.08 (d, J = 7.4Hz, 21H). $^{13}{\rm C}$ NMR (100 MHz, CDCl₃) δ : 196.7, 149.2, 145.4, 144.6, 134.0, 127.6, 121.3, 121.0, 112.3, 87.8, 73.3, 55.7, 31.6, 30.4, 18.0 (6 \times C), 13.0 (3 × C). Minor diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 6.92–6.86 (m, 1H), 6.75–6.70 (m, 3H), 6.13–6.07 (m, 1H), 5.64– 5.60 (m, 1H), 4.00 (dd, J = 8.8, 3.8 Hz, 1H), 3.76 (s, 3H), 2.78-2.54(m, 2H), 2.23-2.15 (m, 1H), 2.02-1.90 (m, 1H). 1.33-1.18 (m, 3H), 1.08 (d, J = 7.4 Hz, 21H). ¹³C NMR (100 MHz, CDCl₃) δ : 196.4, 149.3, 148.0, 145.5, 133.7, 128.9, 121.3, 121.0, 112.3, 91.1, 77.8, 55.7, 32.3. 30.4, 18.0 (6 \times C), 13.0 (3 \times C). HRMS (Cl⁺): m/z calcd for C₂₃H₃₆O₅Si [M]⁺ 420.2332, found 420.2333.

6-Hydroxy-2-(4-((triisopropylsilyl)oxy)phenethyl)-2H-pyran-3(6H)-one (2n). Yellowish oil (dr 2:1; method A, 174 mg, 89%; method B, 156 mg, 80%; method C, 152.3 mg, 78%). IR (neat, cm⁻¹): 3387, 2965, 2945, 2880, 2868, 1683, 1466, 1262, 1130, 1013. Major

diastereomer. ¹H NMR (400 MHz, CDCl₃) δ : 7.05 (d, J = 8.2 Hz, 2H), 6.89 (td, J = 10.5, 2.4 Hz, 1H), 6.79 (dd, J = 8.2, 1.7 Hz, 2H), 6.14–6.07 (m, 1H), 5.64–5.60 (m, 1H), 4.54 (dd, J = 8.4, 3.7 Hz, 1H), 2.81–2.56 (m, 2H), 2.28–2.20 (m, 1H), 2.01–1.94 (m, 1H), 1.26–1.23 (m, 3H), 1.10–1.08 (m, 21H). ¹³C NMR (100 MHz, CDCl₃) δ : 196.9, 154.3, 148.0, 133.8, 129.5, 127.6, 119.9, 87.7, 73.3, 30.3, 18.0 (6 × C), 12.8 (3 × C). Minor diastereomer. ¹H NMR (400 MHz, CDCl₃) δ : 7.05 (d, J = 8.2 Hz, 2H), 6.89 (td, J = 10.5, 2.4 Hz, 1H), 6.79 (dd, J = 8.2, 1.7 Hz, 2H), 6.14–6.07 (m, 1H), 4.01 (dd, J = 8.9, 3.7 Hz, 1H), 2.81–2.56 (m, 2H), 2.28–2.20 (m, 1H), 2.01–1.94 (m, 1H), 1.26–1.23 (m, 3H), 1.10–1.08 (m, 21H). ¹³C NMR (100 MHz, CDCl₃) δ : 196.5, 154.3, 144.7, 133.5, 129.5, 128.8, 119.9, 91.0, 77.8, 31.6, 18.0 (6 × C), 12.8 (3 × C). HRMS (Cl⁺): m/z calcd for $C_{22}H_{34}O_{4}$ Si [M]⁺ 390.2226, found 390.2225.

6-Hydroxy-2-(3,4,5-trimethoxyphenethyl)-2H-pyran-3(6H)-one (20). Yellowish oil (dr 5:3; method A, 114 mg, 74%; method B, 120.2 mg, 78%; method C, 111 mg, 72%). IR (neat, cm $^{-1}$): 3381, 2961, 2948, 2885, 2864, 1689, 1460, 1266, 1135, 1010. Major diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 6.95–6.89 (m, 1H), 6.43 (s, 2H), 6.08 (d, J = 12.0, 1H), 5.67 (s, 1H), 4.57 (dd, J = 8.2, 3.6 Hz, 1H), 3.84 (s, 6H), 3.81 (s, 3H), 2.77–2.62 (m, 2H), 2.27–2.18 (m, 1H), 2.09–1.95 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ: 196.6, 153.2, 144.6, 137.4, 136.2, 127.7, 105.6, 87.8, 77.4, 73.2, 56.2, 31.6, 31.5. Minor diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 6.95–6.89 (m, 1H), 6.43 (s, 2H), 6.08 (d, J = 12.0, 1H), 5.67 (s, 1H), 4.06 (dd, J = 8.6, 3.8 Hz, 1H), 3.84 (s, 6H), 3.81 (s, 3H), 2.77–2.62 (m, 2H), 2.27–2.18 (m, 1H), 2.09–1.95 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ: 196.2, 153.3, 148.0, 137.2, 136.2, 128.9, 105.6, 91.1, 77.9, 61.0, 56.2, 32.4, 31.6. HRMS (Cl $^+$): m/z calcd for C₁₆H₂₀O₆ [M] $^+$ 308.1260, found 308.1260.

6-Hydroxy-2-isopropyl-6-methyl-2H-pyran-3(6H)-one (**2p**). ³² Yellowish oil (dr 6:1; method A, 73.2 mg, 86%; method B, 78.3 mg, 92%; method C, 71.5 mg, 84%). Major diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 6.80 (dd, J = 8.0, 4.0 Hz, 1H), 5.97 (dd, J = 10.2, 2.5 Hz, 1H), 4.32 (s, 1H), 3.36 (brs, 1H), 2.41–2.36 (m, 1H), 1.61 (s, 3H), 1.00 (dd, J = 6.9, 2.4 Hz, 3H), 0.82 (dd, J = 6.8, 2.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 197.5, 148.4, 127.0, 92.6, 78.4, 28.8, 28.7, 19.1, 16.1. Minor diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 6.84 (d, J = 8.0 Hz, 1H), 5.97 (dd, J = 10.2, 2.5 Hz, 1H), 3.94 (t, J = 3.1 Hz, 1H), 3.36 (brs, 1H), 2.41–2.36 (m, 1H), 1.56 (s, 3H), 1.00 (dd, J = 6.9, 2.4 Hz, 3H), 0.90 (dd, J = 6.9, 2.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 196.7, 151.2, 126.9, 94.7, 82.1, 29.5, 24.0, 19.0, 16.8.

6-((Benzyloxy)methyl)-6-hydroxy-2-(2-((triisopropylsilyl))oxy)-ethyl)-2H-pyran-3(6H)-one (2q). ^{26b} Yellowish oil (dr 7:1; method A, 180.2 mg, 83%; method B, 171.7 mg, 79%; method C, 197.8 mg, 91%). Major diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 7.39–7.30 (m, 5H), 6.78 (d, J = 10.3 Hz, 1H), 6.10 (d, J = 10.2 Hz, 1H), 4.79 (dd, J = 8.5, 3.7 Hz, 1H), 4.74 (d, J = 11.9 Hz, 1H), 4.64 (d, J = 12.1 Hz, 1H), 3.85 (dd, J = 7.3, 5.3 Hz, 2H), 3.61 (q, J = 10.3 Hz, 2H), 2.29 (dtd, J = 14.3, 7.3, 3.8 Hz, 1H), 1.86 (ddd, J = 14.0, 8.6, 5.2 Hz, 1H), 1.12–1.01 (m, 21H). ¹³C NMR (100 MHz, CDCl₃) δ: 197.0, 144.8, 137.5, 128.6, 128.1, 127.9, 127.5, 93.0, 74.5, 74.2, 71.4, 59.0, 33.1, 18.1 (6 × C), 12.1 (3 × C).

6-Hydroxy-6-methyl-2H-pyran-3(6H)-one (2r). ³² Yellowish oil (method A, S4.5 mg, 85%; method B, S0 mg, 78%; method C, 51.3 mg, 80%). ¹H NMR (400 MHz, CDCl₃) δ: 6.87 (d, J = 10.3 Hz, 1H), 6.06 (d, J = 10.3 Hz, 1H), 4.56 (d, J = 17.0 Hz, 1H), 4.11 (d, J = 16.9 Hz, 1H), 1.64 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 195.0, 149.0, 126.6, 93.0, 66.7, 28.1.

6-Hydroxy-2-(4-methoxyphenyl)-2H-pyran-3(6H)-one (2s). ³⁷ Yellowish oil (dr 2:1; method A, 102.4 mg, 93%; method B, 90.3 mg, 82%; method C, 93.6 mg, 85%). Major diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 7.28–7.22 (m, 3H), 6.98–6.89 (m, 3H), 6.19 (d, J = 10.3 Hz, 1H), 5.66 (d, J = 3.3 Hz, 1H), 5.52 (s, 1H), 3.79 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 195.2, 159.9, 145.1, 129.5, 127.9, 114.1, 88.1, 76.8, 55.4. Minor diastereomer. ¹H NMR (400 MHz, CDCl₃) δ: 7.28–7.22 (m, 3H), 6.98–6.89 (m, 3H), 6.24 (d, J = 10.3 Hz, 1H), 5.73 (d, J = 1.6 Hz, 1H), 5.02 (s, 1H), 3.79 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 194.8, 159.9, 148.5, 129.4, 127.6, 114.0, 91.6, 80.9, 55.4.

(2*R*,6*S*)-6-Hydroxy-2-methyl-1-tosyl-1,6-dihydropyridin-3(2*H*)-one (2*t*). ³⁸ Colorless oil (method A, 121 mg, 86%; method B, 97 mg, 69%; method C, 104.1 mg, 74%). ¹H NMR (400 MHz, C_6D_6) δ: 7.56 (d, J = 8.0 Hz, 2H), 6.70 (d, J = 7.9 Hz, 2H), 6.21 (dd, J = 10.3, 4.5 Hz, 1H), 5.83 (d, J = 4.6 Hz, 1H), 5.56 (d, J = 10.3 Hz, 1H), 4.60 (q, J = 7.3 Hz, 1H), 1.81 (s, 3H), 1.61 (d, J = 7.2 Hz, 3H). ¹³C NMR (100 MHz, C_6D_6) δ: 194.8, 143.8, 143.8, 137.7, 130.1, 127.1, 125.81, 73.8, 57.6, 21.9, 21.1.

Preparation of (2R.6S)-6-Phenyl-2-(2-((triisopropylsilyl)oxy)ethyl)-2H-pyran-3(6H)-one (3). To a stirred solution of the crude product 2b (5.23 g, 16.7 mmol) in CH_2Cl_2 (40 mL) were added acetic anhydride (Ac₂O, 2.56 g, 25.1 mmol), Et₃N (2.54 g, 25.1 mmol) and 4-dimethylaminopyridine (DMAP, 0.41 g, 3.34 mmol) at 0 °C. The reaction mixture was stirred at room temperature overnight. The reaction was guenched by addition of saturated aqueous NH₄Cl (20 mL) and extracted with CH₂Cl₂ (3 × 20 mL). The combined organic fractions were washed with brine, dried over Na2SO4, filtered and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (EtOAc/hexane =1/8-1/3) to afford the desired acetylated product (5.72 g, 16.0 mmol, 96% yield) as a 5:4 diastereomeric mixture. $\left[\alpha_D^{20}\right] = +16.8$ (c 1.1, MeOH). IR (neat, cm⁻¹): 2944, 2867, 1757, 1699, 1464, 1370, 1222, 1172, 1103, 998, 932. Major diastereomer: ¹H NMR (400 MHz, CDCl₃) δ : 6.86–6.81 (m, 1H), 6.47 (d, I = 3.2 Hz, 1H), 6.22–6.18 (m, 1H), 4.72 (dd, J = 8.4, 3.6 Hz, 1H), 3.83-3.78 (m, 2H), 2.27-2.23 (m, 1H), 2.10 (s, 3H), 1.83-1.78 (m, 1H), 1.06-1.01 (m, 21H). ^{13}C NMR (100 MHz, CDCl₃) δ : 195.9, 169.6, 143.0, 128.8, 88.0, 76.2, 58.6, 35.9, 21.1, 18.0 (6 \times C), 12.0 (3 \times C). Minor diastereomer: 1 H NMR (400 MHz, CDCl₃) δ : 6.86–6.81 (m, 1H), 6.53 (s, 1H), 6.22– 6.18 (m, 1H), 4.47 (dd, I = 9.6, 4.0 Hz, 1H), 3.83–3.78 (m, 2H), 2.27-2.23 (m, 1H), 2.07 (s, 3H), 2.00-1.96 (m, 1H), 1.06-1.01 (m, 21H). ¹³C NMR (100 MHz, CDCl₃) δ : 195.8, 169.2, 141.2, 128.7, 87.1, 72.5, 58.3, 33.1, 21.0, 18.0 (6 × C), 12.0 (3 × C). HRMS (TOF, CI⁺) m/z calcd for $C_{18}H_{33}O_5Si$ [M + H]⁺ 357.2097, found 357.2097. To a stirred solution of the substrate above (3.04 g, 9.67 mmol) in acetic acid (20 mL) was added activated Zn powder (3.09 g, 48.34 mmol) at room temperature. The reaction mixture was stirred for 1.5 h. The reaction was then quenched by addition of saturated aqueous K_2CO_3 (30 mL) and extracted with CH_2Cl_2 (3 × 20 mL). The combined organic fractions were washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The crude product was used for next step without further purification. To a stirred solution of the crude product obtained above in CH₃CN (10 mL) were added Pd₂(dba)₃ (0.70 g, 0.76 mmol), NaOAc (1.88 g, 22.9 mmol) and phenyldiazonium salt (2.92 g, 15.2 mmol) at room temperature. The reaction mixture was stirred for 4 h at room temperature. The reaction was quenched by addition of saturated aqueous NH₄Cl (30 mL) and extracted with EtOAc (3 \times 20 mL). The combined organic fractions were washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (EtOAc/hexane =1/10-1/4) to afford the desired product (-)-3 (2.61 g, 6.96 mmol, 72% yield over two steps) as a reddish oil. $\left[\alpha_{\rm D}^{20}\right] = -18.6$ (c 1.0, MeOH). IR (neat, cm⁻¹): 3032, 2943, 2866, 1693, 1463, 1384, 1259, 1102, 1055, 998, 883. ¹H NMR (400 MHz, CDCl₃) δ : 7.39–7.35 (m, 5H), 7.16 (dd, J = 10.4, 3.2, 1H), 6.24 (dd, J= 10.4, 1.9, 1H), 5.50 (brs, 1H), 4.36 (dd, *J* = 9.2, 3.9, 1H), 3.82 (dd, *J* = 7.4, 5.1, 2H), 2.12 (dtd, J = 14.5, 7.4, 4.0, 1H), 1.93 (ddt, J = 14.2, 14.2, 14.2)9.7, 5.0, 1H), 0.98 (d, J = 4.4, 21H). ¹³C NMR (100 MHz, CDCl₃) δ : 196.9, 149.1, 136.8, 128.8 (2 × C), 128.7, 128.0 (2 × C), 126.5, 73.6, 72.7, 59.1, 33.0, 18.1 (4 \times C), 17.8 (2 \times C), 12.4 (2 \times C), 12.0. HRMS (TOF, Cl⁺) m/z calcd for $C_{22}H_{35}O_3Si$ [M + H]⁺ 375.2355, found 375,2360.

Preparation of 1,6-Dioxaspiro[4.5]dec-9-en-8-one (4). To a stirred solution of furfuryl alcohol 1u (102 mg, 0.74 mmol) in MeCN (4 mL) and H_2O (0.2 mL) were added KBr (4.4 mg, 0.037 mmol), NaHCO₃ (31 mg, 0.37 mmol), and Oxone (0.55 g, 0.89 mmol) at 0 °C. After completion of the addition, the reaction was allowed to stir at 0 °C for 30 min. After completion of the Achmatowicz reaction as monitored by TLC, 10-camphorsulfonic acid (CSA, 150 mg, 0.74

mmol) was added, and the reaction mixture was allowed to warm to room temperature for 1 h. The reaction was then quenched by addition of saturated aqueous NaHCO₃ (1 mL) with EtOAc (3 × 1 mL). The combined organic fractions were washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (EtOAc/hexane = 1/10-1/5) to afford the desired product 4^{27} (81 mg, 0.53 mmol, 71% yield over two steps) as a yellowish oil. ¹H NMR (400 MHz, CDCl₃) δ : 6.77 (d, J = 10.2 Hz, 1H), 6.11 (d, J = 10.2 Hz, 1H), 4.49 (d, J = 16.8 Hz, 1H), 4.11–3.98 (m, 3H), 2.27–2.16 (m, 2H), 2.11–1.93 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 195.6, 147.6, 128.0, 102.3, 68.9, 67.1, 37.8, 24.8.

Preparation of (2S)-2-(2-Bromo-4-methoxy-5-((triisopropylsilyl)oxy)phenethyl)-6-hydroxy-2H-pyran-3(6H)one (5). To a stirred solution of furfuryl alcohol 1m (104 mg, 0.27 mmol) in THF (1 mL) were added KBr (0.1 M in H₂O, 0.13 mL, 0.013 mmol), NaHCO₃ (1 M in H₂O, 0.13 mL, 0.13 mmol), and Oxone (0.20 g, 0.32 mmol) at 0 °C. After completion of the addition, the reaction was allowed to stir at 0 °C for 30 min. After completion of the Achmatowicz reaction as monitored by TLC, KBr (32 mg, 0.27 mmol) and Oxone (0.17 g, 0.27 mmol) were added, and the reaction mixture was allowed to warm to room temperature for 1 h. The reaction was then quenched by addition of saturated aqueous NaHCO₃ (1 mL) and EtOAc (3 \times 1 mL). The combined organic fractions were washed with brine, dried over Na2SO4, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (EtOAc/hexane = 1/10-1/5) to afford the desired product 5 (115 mg, 0.23 mmol, 85% yield) as a yellowish oil (dr = 2/1). IR (neat, cm⁻¹): 3395, 2964, 2937, 2883, 2860, 1684, 1452, 1264, 1130, 1014. Major diastereomer. ¹H NMR (400 MHz, CDCl₃) δ : 6.95 (s, 1H), 6.94–6.85 (m, 1H), 6.76 (s, 1H), 6.11 (dd, J = 15.1, 10.2 Hz, 1H), 5.74-5.59 (m, 1H), 4.55 (dd, J = 8.3, 3.7 Hz, 1H), 3.75 (s, 3H), 2.82–2.68 (m, 2H), 2.26– 2.10 (m, 1H), 1.94 (tdd, *J* = 17.0, 8.8, 4.6 Hz, 1H), 1.22 (ddd, *J* = 14.7, 9.6, 7.1 Hz, 4H), 1.06 (d, J = 7.4 Hz, 18H). ¹³C NMR (100 MHz, CDCl₃) δ : 196.5, 150.0, 144.6, 132.7, 127.6, 122.0, 116.3, 114.6, 87.8, 73.4, 55.8, 30.9, 30.1, 18.0 (6 \times C), 13.0 (3 \times C). Minor diastereomer. ¹H NMR (400 MHz, CDCl₃) δ : 6.95 (s, 1H), 6.94–6.85 (m, 1H), 6.76 (s, 1H), 6.11 (dd, J = 15.1, 10.2 Hz, 1H), 5.74-5.59 (m, 1H), 4.03 (dd, J = 8.8, 3.8 Hz, 1H), 3.75 (s, 3H), 2.82 - 2.68 (m, 2H), 2.26 -2.10 (m, 1H), 1.94 (tdd, *J* = 17.0, 8.8, 4.6 Hz, 1H), 1.22 (ddd, *J* = 14.7, 9.6, 7.1 Hz, 4H), 1.06 (d, J = 7.4 Hz, 18H). ¹³C NMR (100 MHz, $CDCl_3$) δ : 196.1, 149.9, 147.9, 145.0, 132.5, 128.8, 122.1, 116.4, 114.7, 91.0, 77.8, 55.8, 30.9, 18.0 (6 \times C), 13.0 (3 \times C). HRMS (Cl⁺): m/zcalcd for C₂₃H₃₅BrO₅Si [M]⁺ 498.1437, found 498.1431.

Preparation of 2-Isopropyl-2H-pyran-3(4H)-one (6). To a stirred solution of furfuryl alcohol 1a (0.1 g, 0.71 mmol) in THF (4 mL) at 0 °C were added KBr (0.1 M in H₂O, 0.36 mL, 0.036 mmol), NaHCO₃ (1 M in H₂O, 0.36 mL, 0.036 mmol), and Oxone (0.52 g, 0.85 mmol) at 0 °C. The reaction was allowed to stir for 30 min. The reaction was then quenched by addition of saturated aqueous NaHCO₃ (1 mL) and extracted with EtOAc (3 \times 3 mL). The combined organic fractions were washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The crude product 2a was used for the next step without further purification. To a stirred solution of the crude product 2a in acetic acid (2 mL) was added activated Zn powder (0.23 g, 3.55 mmol) at room temperature. The reaction mixture was stirred for 1.5 h. The reaction was then quenched by addition of saturated aqueous K₂CO₃ (5 mL) and extracted with CH_2Cl_2 (3 × 4 mL). The combined organic fractions were washed with brine, dried over Na2SO4, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (EtOAc/hexane = 1/10-1/5) to afford the desired product 6^{25} (51 mg, 0.36 mmol, 51% yield over two steps) as a colorless oil. ¹H NMR (400 MHz, CDCl₃) δ : 6.29–6.09 (m, 1H), 4.34 (dt, J = 5.4, 3.6 Hz, 1H), 3.67 (d, J = 4.6 Hz, 1H), 2.54-2.28 (m, 2H), 2.21 (dq, J = 13.4, 6.6 Hz, 1H), 0.89 (dd, J = 15.8, 6.8 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ: 205.0, 143.6, 98.1, 86.0, 35.0, 29.5, 18.8, 17.2.

Preparation of 6-Isopropyl-5-oxo-5,6-dihydro-2H-pyran-2-yl acetate (7a). To a stirred solution of crude product 2a [obtained from furfuryl alcohol 1a (0.1 g, 0.71 mmol) without purification] in CH₂Cl₂ (20 mL) were added acetic anhydride (Ac₂O, 0.11 g, 1.07 mmol), Et₃N (0.11 g, 1.07 mmol), and 4-(dimethylamino)pyridine (DMAP, 6.1 mg, 0.04 mmol) at 0 °C. The reaction mixture was stirred at room temperature overnight. The reaction was quenched by addition of saturated aqueous NH₄Cl (4 mL) and extracted with CH_2Cl_2 (3 × 2 mL). The combined organic fractions were washed with brine, dried over Na2SO4, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 4:1) to afford the desired product 7a²⁵ (115 mg, 0.58 mmol) in 82% yield over two steps as a 1.1:1 diastereomeric mixture. Major diastereomer. ¹H NMR (400 MHz, C_6D_6) δ : 6.35 (dd, J = 13.6, 2.7 Hz, 1H), 6.19–6.06 (m, 1H), 5.91-5.73 (m, 1H), 4.18 (d, J = 2.8 Hz, 1H), 2.46 (pd, J = 7.0, 2.8 Hz, 1H), 1.60 (dd, I = 5.2, 1.1 Hz, 3H), 1.04–0.85 (m, 6H). ¹³C NMR (100 MHz, C_6D_6) δ : 194.9, 168.8, 141.4, 128.9, 87.3, 80.1, 30.3, 18.9, 16.2. Minor diastereomer. ¹H NMR (400 MHz, C_6D_6) δ : 6.35 (dd, J =13.6, 2.7 Hz, 1H), 6.19-6.06 (m, 1H), 5.91-5.73 (m, 1H), 3.64 (d, J = 5.6 Hz, 1H), 2.30 (dq, J = 13.3, 6.6 Hz, 1H), 1.60 (dd, J = 5.2, 1.1 Hz, 3H), 1.04–0.85 (m, 6H). 13 C NMR (100 MHz, C_6D_6) δ : 194.4, 168.6, 143.6, 129.3, 88.5, 84.1, 29.1, 20.4, 17.6.

Preparation of tert-Butyl (6-Isopropyl-5-oxo-5,6-dihydro-2H-pyran-2-yl) carbonate (7b). Following the procedure for the synthesis of 7a, $7b^{25}$ (133 mg, 0.52 mmol) (dr = 1.2:1) was obtained from 1a (0.1 g, 0.71 mmol) using (Boc)₂O (0.23 g, 1.07 mmol), Et₃N (0.11 g, 1.07 mmol), and 4-(dimethylamino)pyridine (DMAP, 6.1 mg, 0.04 mmol) in 73% yield over two steps. Major diastereomer: ¹H NMR (400 MHz, C_6D_6) δ : 6.23 (dd, J = 15.0, 2.8 Hz, 1H), 6.13 (ddd, J = 10.2, 3.7, 1.3 Hz, 1H), 5.77 (dd, <math>J = 14.2, 10.2 Hz, 1H), 4.21 (d, <math>J = 14.2, 10.2 Hz2.9 Hz, 1H), 2.40 (dtt, J = 9.8, 7.1, 2.8 Hz, 1H), 1.35–1.26 (m, 9H), 1.00-0.84 (m, 6H). ¹³C NMR (100 MHz, C_6D_6) δ : 194.7, 152.4, 140.7, 129.1, 90.5, 82.5, 79.8, 28.9, 27.6, 18.8, 16.1. Minor diastereomer: ¹H NMR (400 MHz, C_6D_6) δ : 6.23 (dd, J = 15.0, 2.8 Hz, 1H), 6.13 (ddd, J = 10.2, 3.7, 1.3 Hz, 1H), 5.77 (dd, J = 14.2, 10.2Hz, 1H), 3.64 (d, J = 6.6 Hz, 1H), 2.30 (dq, J = 13.5, 6.9 Hz, 1H), 1.35-1.26 (m, 9H), 1.00-0.84 (m, 6H). 13 C NMR (100 MHz, C_6D_6) δ: 194.4, 152.4, 142.5, 129.1, 89.7, 84.3, 82.7, 30.7, 27.6, 18.8, 18.0.

Preparation of 6-Isopropyl-2H-pyran-2,5(6H)-dione (8). To a stirred solution of the crude product 2a [obtained from furfuryl alcohol 1a (0.2 g, 1.43 mmol) without purification] in acetone (10 mL) at 0 °C was added Jones reagent (1.5 mL, 2.9 M) dropwise. After being stirred at 0 °C for 30 min, TLC showed the complete consumption of 2a, and the reaction was quenched by slow addition of i-PrOH (0.7 mL) at 0 °C. The mixture was filtered through a pad of Celite and washed with diethyl ether. The filtrate was washed with brine (2 × 5 mL). The organic layer was dried over MgSO₄, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 4:1) to afford the desired product 8 (183 mg, 1.19 mmol) in 83% yield over two steps. IR (neat, cm⁻¹): 2945, 2922, 2855, 1725, 1693, 1465, 1363, 1267, 1126, 1087, 967. ¹H NMR (400 MHz, CDCl₃) δ : 6.80 (d, J = 10.1 Hz, 1H), 6.68 (d, J = 10.2 Hz, 1H), 4.69 (d, J = 3.4 (d))Hz, 1H), 2.39-2.18 (m, 1H), 0.99 (d, J = 7.1 Hz, 3H), 0.80 (d, J = 7.0Hz, 3H). 13 C NMR (100 MHz, CDCl₃) δ : 193.3, 160.6, 138.7, 135.3, 88.3, 33.1, 18.5, 15.7. HRMS (Cl⁺): m/z calcd for $C_8H_{11}O_3$ [M + H]⁺ 155.0703, found 155.0704.

Preparation of (2R,6R)-6-((Benzyloxy)methyl)-2-((2-((triisopropylsilyl)oxy)ethyl)-2H-pyran-3(6H)-one (10). To a stirred solution of furfuryl alcohol 1q (0.1 g, 0.24 mmol) in THF (1 mL) at 0 °C were added KBr (0.1 M in H₂O, 0.12 mL, 0.012 mmol), NaHCO₃ (1 M in H₂O, 0.12 mL, 0.12 mmol), and Oxone (0.18 g, 0.29 mmol) at 0 °C. The reaction was allowed to stir for 30 min. The reaction was then quenched by addition of saturated aqueous NaHCO₃ (1 mL) and extracted with EtOAc (3 × 3 mL). The combined organic fractions were washed with brine, dried over Na₂SO₄, and concentrated under reduced pressure. The crude product 2q was used for the next step without further purification. To a stirred solution of the crude pyranone product 2q in CH₂Cl₂ (2 mL) at -78

°C under nitrogen atmosphere were added triethylsilane (Et₃SiH, 0.23 mL, 1.44 mmol) and boron trifluoride diethyl etherate (BF₃·Et₂O₄ 0.06 mL, 0.48 mmol) dropwise. The reaction mixture was stirred at -78 °C for 1 h, and then the reaction was quenched by addition of saturated aqueous NaHCO3 (10 mL). The organic layer was collected, and the aqueous layer was extracted with CH_2Cl_2 (3 × 5 mL). The combined organic fractions were washed with brine, dried over MgSO4, and concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel using eluents (ethyl acetate/hexane = 1/20-1/10) to give the dihydropyranone product 10²⁶ (80 mg, 0.19 mmol, 80% yield over two steps) as a colorless oil. ¹H NMR (400 MHz, CDCl₃) δ : 7.39–7.27 (m, 5H), 7.06 (dd, J =10.3, 1.5 Hz, 1H), 6.16 (dd, J = 10.4, 2.4 Hz, 1H), 4.70-4.55 (m, 2H), 4.50 (dg, I = 6.4, 2.8 Hz, 1H), 4.25 (dt, I = 9.1, 2.6 Hz, 1H), 3.96 - 3.79(m, 2H), 3.71 (dd, J = 10.0, 5.5 Hz, 1H), 3.59 (dd, J = 10.0, 5.9 Hz, 1H), 2.34 (dddd, J = 12.9, 9.4, 6.2, 3.4 Hz, 1H), 1.78 (ddt, J = 13.8, 9.0, 4.5 Hz, 1H), 1.05 (d, J = 5.0 Hz, 21H). ¹³C NMR (100 MHz, CDCl₃) δ : 196.9, 148.7, 137.9, 128.6, 128.0, 127.9, 127.8, 77.2, 73.8, 73.7, 71.2, 58.9, 33.1, 18.1 (6 \times C), 12.1 (3 \times C).

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.6b00469.

¹H and ¹³C NMR spectra of new compounds and HPLC chromatograms of compounds **1b** and **3** (PDF)

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Notes

The authors declare no competing financial interest.

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